

## Abstract

**Background:** Chronic arsenic exposure results in multi-organ cancers, skin being the major target. Arsenic is a clastogen and exposure to arsenic leads to double strand break (DSB) accumulation, an important step in carcinogenesis<sup>1,2</sup>. The Ataxia-telangiectasia mutated (ATM) protein is an apical kinase that is required for high-fidelity DSB DNA damage response recruitment. Following successful DSB repair, phosphatases (PP2A and PP5) dephosphorylate ATM.

**Hypothesis:** Arsenic exposure suppresses DSB repair response leading to accumulation of double-strand DNA breaks. To examine this hypothesis, the DSB repair response was analyzed in immortalized human keratinocytes (HaCaT and Ker-CT cells) chronically and acutely exposed to sodium arsenite (iAs<sup>3+</sup>).

**Materials and Methods:** HaCaT and Ker-CT cells were chronically exposed to iAs<sup>3+</sup> (0.1 μM; 7 and 8 weeks respectively). HaCaT cells were also acutely exposed to increasing doses of iAs<sup>3+</sup> (0-5 μM; 24 h). Cell lysates were collected and subjected to immunoblot analysis for ATM (activated and total), its downstream protein targets (CHK2, TP53, CDKN1A; activated and total) as well as phosphatases (PP2ACA, PPP2R2C, PP5). Densitometric data was analyzed employing unpaired student's two-tailed t-test with a Welch's correction (chronic exposure) or One-way ANOVA followed by Tukey's multiple comparisons post-hoc test (acute exposure).

**Results:** ATM phosphorylation was significantly reduced upon chronic iAs<sup>3+</sup> exposure in both cell lines. Such exposure also led to induction of PPP2R2C and PP5C (mRNA and protein) in HaCaT cells but suppression of PPP2R2C and PP5C (protein) in Ker-CT cells. Acute iAs<sup>3+</sup> exposure in HaCaT cells induced PPP2R2C protein expression.

**Conclusions:** Chronic iAs<sup>3+</sup> exposure suppressed ATM activation irrespective of the cell line tested. Suppressed ATM activation may lead to an impaired DSB repair response that acts as a harbinger for iAs<sup>3+</sup>-induced skin carcinogenesis. However, the phosphatase expression profile is opposite in the two cell lines, suggesting that ATM activation might be modulated by upstream regulators rather than phosphatases exclusively.

## Background

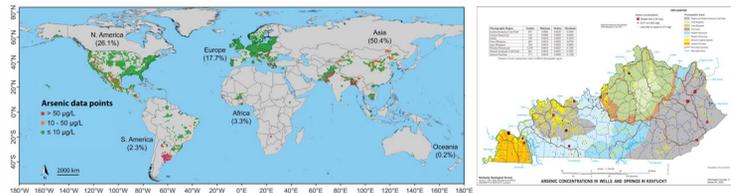


Figure 1. Arsenic contamination in drinking water worldwide and Kentucky<sup>3,4</sup>.

○ Arsenic exposure occurs primarily through contaminated drinking water that exceeds the World Health Organization (WHO) and Environmental Protection Agency (EPA) maximum contaminant level concentration (10 ug/L) (Figure 1)

○ Chronic exposure to low doses of arsenic induces cardiovascular and pulmonary disease, diabetes, and multi-organ cancers, including skin cancer (Figure 2)

○ Arsenic is a clastogen – leads to accumulation of chromosomal aberrations

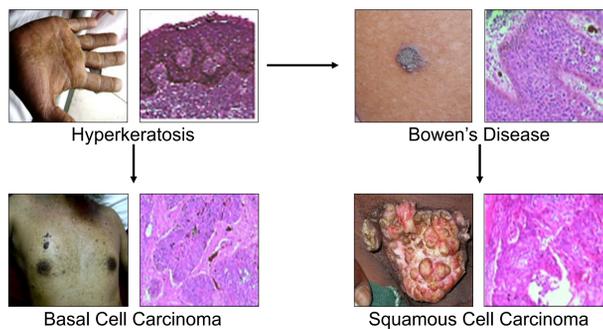


Figure 2. Chronic arsenic exposure can lead to the development of skin cancer<sup>5</sup>.

## Background

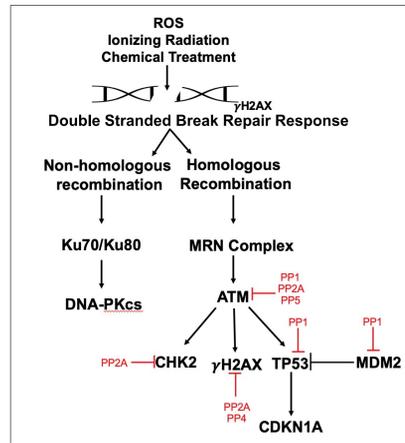


Figure 3A. Two pathways involved in the DSB repair response

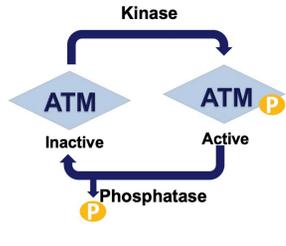


Figure 3B. Catalytic regulation of ATM activation

## Hypothesis and Aims

**Hypothesis:** Arsenic exposure suppresses the DSB repair response leading to accumulation of double-strand DNA breaks.

**Aims:**

- Determine whether the double-strand DNA repair response is perturbed in cells chronically or acutely treated with toxicologically relevant concentrations of iAs<sup>3+</sup>.
- Assess whether commonalities exist between double-strand DNA repair responses across keratinocyte cell lines as a result of chronic iAs<sup>3+</sup> exposure.

## Materials & Methods

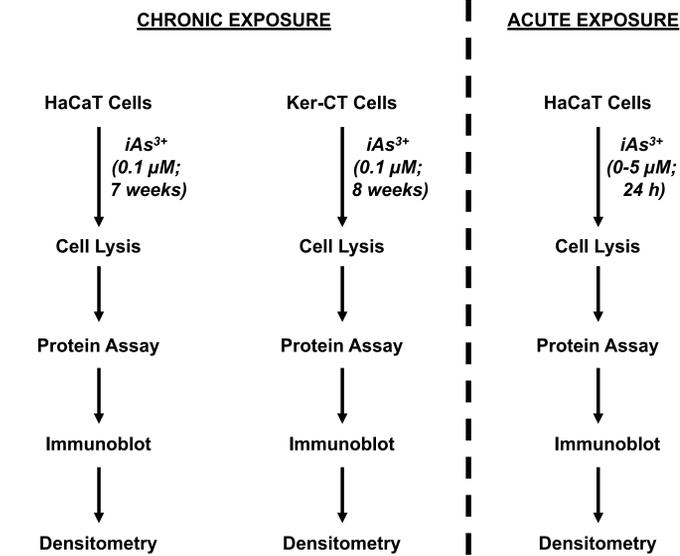


Figure 4A. Experimental strategy for chronic iAs<sup>3+</sup> exposure in HaCaT and Ker-CT cells

Figure 4B. Experimental strategy for acute iAs<sup>3+</sup> exposure in HaCaT cells

## Results

**ATM pathway activation is suppressed by chronic iAs<sup>3+</sup> exposure in HaCaT cells**

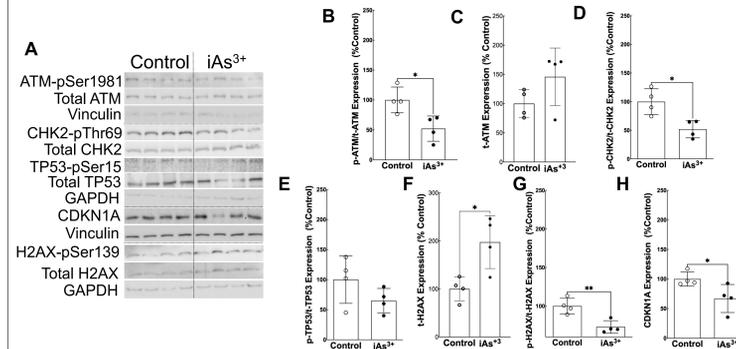


Figure 6 A. Immunoblots for ATM pathway molecules in chronically iAs<sup>3+</sup> exposed HaCaT cells. B-H. Densitometric analysis for activated ATM (B), total ATM (C), activated CHK2 (D), activated TP53 (E), total H2AX (F), activated H2AX (G) and CDKN1A (H). \*p<0.05, \*\*p<0.01 by unpaired, two-tailed t-test with Welch's correction.

**Chronic iAs<sup>3+</sup> exposure induces expression of phosphatases in HaCaT cells**

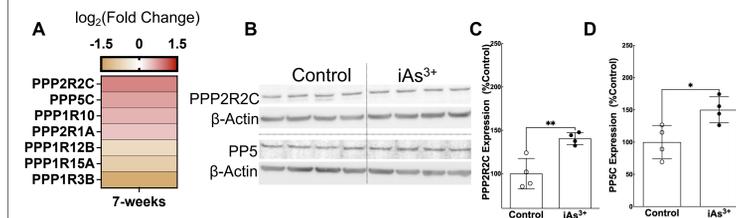


Figure 7A. Modulation of mRNA expression of phosphatases by chronic iAs<sup>3+</sup> exposure. B. Immunoblots for selected phosphatases C-D. Densitometric analysis for PPP2R2C (C) and PP5 (D). \*p<0.05, \*\*p<0.01 by unpaired, two-tailed t-test with Welch's correction.

**Chronic iAs<sup>3+</sup> exposure suppressed ATM activation but induces phosphatase expression in Ker-CT cells**

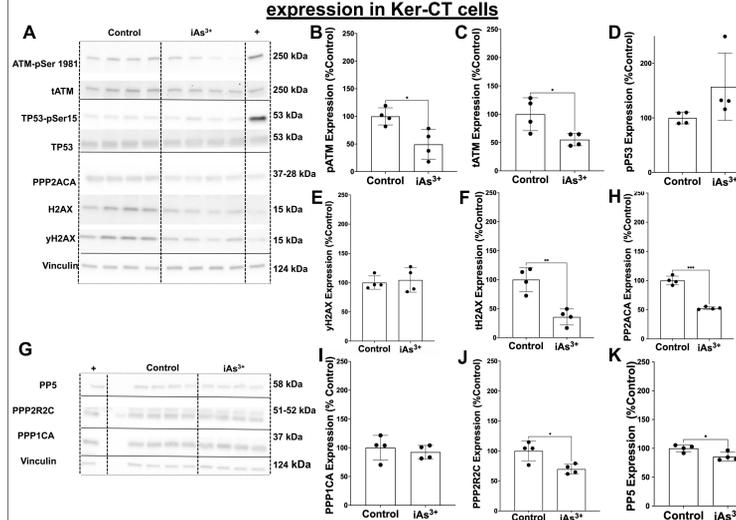


Figure 8 A. Immunoblots for ATM pathway molecules in chronically iAs<sup>3+</sup> exposed Ker-CT cells. B-F. Densitometric analysis for activated ATM (B), total ATM (C), activated TP53 (D), activated H2AX (E) and total H2AX (F). G. Immunoblots for selected phosphatases. H-K. Densitometric analysis for PPP2ACA (H), PPP1CA (I), PPP2R2C (J) and PP5 (K). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 by unpaired, two-tailed t-test with Welch's correction (Panels B-F and H-K).

## Results

**Acute iAs<sup>3+</sup> exposure does not alter ATM activation or phosphatase expression in HaCaT cells**

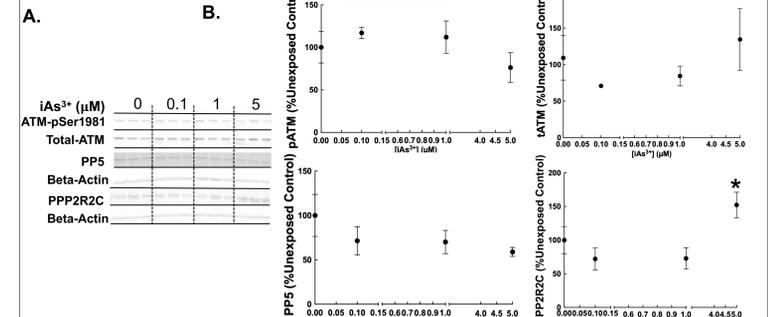


Figure 9A. Immunoblots for ATM (phosphorylated and total) and selected phosphatases in HaCaT cells exposed acutely to increasing doses of iAs<sup>3+</sup>. B-E. Densitometric analysis for activated ATM (B), total ATM (C), PP5 (D) and PPP2R2C (E). \*p<0.05 for One-way ANOVA with Tukey's multiple comparisons post-hoc test.

## Conclusions

**Reduced ATM activation due to chronic iAs<sup>3+</sup> exposure is a common mechanism across two keratinocyte models**

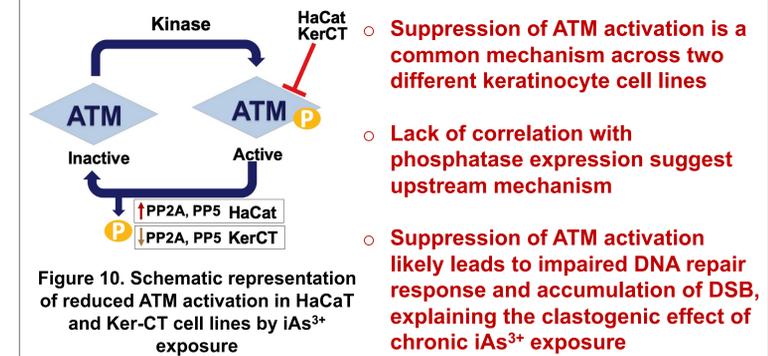


Figure 10. Schematic representation of reduced ATM activation in HaCaT and Ker-CT cell lines by iAs<sup>3+</sup> exposure

## Future Directions

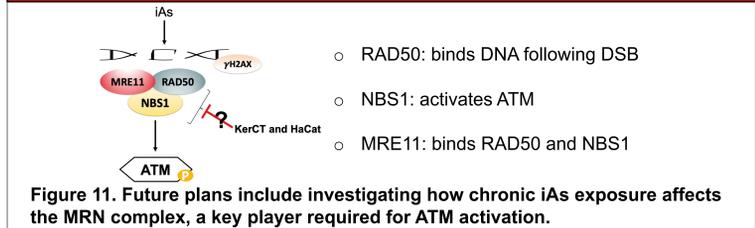


Figure 11. Future plans include investigating how chronic iAs exposure affects the MRN complex, a key player required for ATM activation.

## References

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5. Al-Eryani L, et al. miRNA expression profiles of premalignant and malignant arsenic-induced skin lesions. *PLoS One*. 2018; 13(8):e0202579.

## Acknowledgements

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