

CURRICULUM VITAE

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EDUCATION:

1991 Diploma in Biology
Christian-Albrechts-University of Kiel, Germany

1994 Ph.D. in Molecular Biology
Medical University of Luebeck, Germany

PROFESSIONAL EXPERIENCE

09/2015 – current
37.5 hours/week University Biological Safety Officer, Adjunct Assistant Professor,
School of Public Health and Information Sciences Department of
Environmental Health and Safety, University of Louisville, Louisville,
KY

Responsibilities:

Directs and maintains a comprehensive biological safety program and provides guidance to investigators on the safe use of biohazardous agents in research and clinical activities; ensures compliance with all applicable federal, state and local regulations as well as practices prescribed by the CDC and NIH; serves as the administrator for the IBC, and oversees registrations and authorizations for the use of biohazardous materials; serves as the Responsible Official for the Select Agent Program.

Essential Functions and Duties:

1. Assures compliance of the all aspects of biosafety program with all federal, state, and local regulations.

2. Consults with laboratories on the prevention of occupational exposure to hazardous agents.
3. Measures, audits, and evaluates the effectiveness of hazard control and hazard control programs.
4. Directs the day-to-day operations of the Biosafety Office.
5. Administers the IBC (which oversees use of synthetic and recombinant nucleic acids, RG2 and RG3 organisms, Select Agents and Toxins, human and primate materials).
6. Provides subject matter expertise to the IACUC and IRB (including human gene transfer trials; rDNA use in laboratory, animal, plant, and Select Agents research)
7. Conducts biological safety training (BSL-1 through BSL-3).
8. Updates training on transport of Division 6.2 dangerous goods
 - a. Inspects laboratories to assess biosafety practices of research personnel.
 - b. Investigates spills and incidents.
 - c. Acts as Responsible Official for the Regional Biocontainment Laboratory
9. Accesses staffing needs within the biosafety group and directs and evaluates 3 biosafety/laboratory safety professionals.

Accomplishments:

1. Assessed staffing needs for the biosafety group and proposed/implemented reorganization of the group by creating positions of Associate Biosafety Officer (internal promotion), Laboratory Safety Training Specialist (new) and Laboratory assessments Specialist (new); hiring for the last two positions was completed in June 2016.
2. Developed and implemented institutional procedures for Dual Use Research of Concerns.
3. Revised IBC bylaws to accurately reflect institutional procedures and policies.
4. Developed and implemented IBC procedure manual to formalize and standardize IBC processes.
5. Reviews every month 5-10 IBC protocols and 5-10 animal protocols describing the use of biological materials and hazardous chemical agents and advice on best biosafety practices.
6. Revised Biosafety Manual to incorporate current best biosafety practices.
7. Revised the laboratory safety training program and laboratory inspection program to incorporate adult learning strategies.
8. Prepared for and completed renewal inspection of the U of L Select Agent facility in fall 2016 as well as participated in an unannounced Verification Inspection.
9. Involved as trainer in the Duke Infectious Disease Response Training Program

02/2010 – 09/2015
40 hours/week

Biological Safety Officer, Director of A/BSL-3/Select Agent Facility, Responsible Official for Select Agent Program, Assistant Professor in the Department of Molecular Virology & Microbiology, Department of Environmental Safety, Baylor College of Medicine, Houston, TX

Responsibilities:

Directs and manages the personnel and resources of the BCM Biological Safety Program to achieve its mission, goals and objectives; serves as the institution's Responsible Official (RO) for the Select Agent Program and as the Facility Manager/Director of BCM's BSL-3/3E Facility.

Essential Functions and Duties:

1. Assures compliance with applicable safety and health regulations.
2. Consults with laboratories on the prevention of occupational exposure to hazardous agents.
3. Measures, audits, and evaluates the effectiveness of hazard control and hazard control programs.
4. Directs the day-to-day operations of the Biosafety Office.
5. Directs A/BSL-3/Select Agent Facility.
6. Provides subject matter expertise to the IACUC and IBC (including human gene transfer trials; rDNA use in laboratory, animal, plant, and Select Agents research).
7. Chairs Environmental Safety Committee which reviews the use of hazardous biological (other than rDNA including RG-1, RG-2, and RG-3 organisms) and chemical agents.
8. Conducts biological safety training (BSL-1 through BSL-3).
9. Develops and updates training on transport of Division 6.2 dangerous goods
10. Inspects laboratories to assess biosafety practices of research personnel.
11. Investigates spills and incidents.
12. Acts as Responsible Official for the BCM Select Agent Program.
13. Serves as back up for the Director of Environmental Safety.
14. Supervises Environmental Safety employees.

Accomplishments:

1. Participated in 4 CDC inspections of the BCM Select Agent Program resulting in securing a 3 year approval of the Select Agent Registration.

2. Developed and implemented all changes to the Select Agent Program to comply with major changes to the Select Agent Regulation in 2013.
3. Developed and provided all annually required training to Select Agent User.
4. Assisted Principal investigators in developing containment procedures for vector borne disease (ticks, mosquitoes).
5. Assisted PI in retro fitting standard BSL-3 space to BSL-3 space for use with infected mosquitoes.
6. Directed and participated in annual re-certification of the BCM Select Agent A/BSL-3 facility.
7. Developed and implemented a risk-based laboratory inspection program.
8. Reviewed every month 10 - 15 IBC protocols and 10 - 15 animal protocols describing the use of biological materials and hazardous chemical agents.
9. Participated in all IACUC animal facility inspection year around.
10. Updated and revised training for shipping biological materials every two years
11. Updated and revised Biosafety Manual in 2013
12. Participated in and successfully resolved two FAA for cause inspection without fines.
13. Participated in 2 AAALAC site visits.
14. Participated in developing BCM response to Ebola outbreak for BCM outpatient clinic and provided educational training on Ebola and hands-on training on donning and doffing PPE to clinical staff.
15. Provided BSL-3 hands-on training to users of the BCM BSL-3 facility.

08/2003 – 01/2010
40 hours/week

Administrator for the Institutional Animal Care and Use Committee
and Institutional Biosafety Committee
Office of Research, Baylor College of Medicine, Houston, Texas

Responsibilities:

1. Directed the day-to-day operations of the IACUC/IBC Office.
2. Supervised Research Subject Protection analysts and Training Specialist and helps them resolve problems, issues, and concerns.
3. Provided administrative support to the Chairs of the IACUC and IBC and the Institutional Veterinarian in the collection and maintenance of animal use data, preparation of correspondence, and investigations of allegations of misconduct or ethics concerns in studies.
4. Performed administrative/scientific review for animal use protocols, modifications, and amendments.

5. Maintained and enhance the electronic animal protocol review system; develop electronic review rDNA review system.
6. Participated in semi-annual program review and facility inspections
7. Prepared and maintains all records of IACUC and IBC activities as prescribed by federal oversight agencies.
8. Ensured timely processing of modification memos and other correspondence to and from investigators.
9. Maintained performance measurement data with reporting to the IACUC and IBC on a scheduled basis.
10. Ensured quality customer service delivery from IACUC/IBC office staff.
11. Coordinated and implement with the Institutional Veterinarian IACUC policies and administrative functions in accordance with regulations and accrediting agency policies and procedures.
12. Acted as liaison between the IACUC and IBC and other institutional safety committees and programs, to ensure compliance with state and federal requirements as well as a timely process of protocol approvals from those committees.
13. Established working relationships with investigators, faculty and staff to ensure a timely and complete protocol review and approval process.
14. Provided education on research ethics and regulatory requirements to the research community.

Accomplishments:

1. Organized the newly formed IACUC office in 2003 and re-organized IBC office in 2005.
2. Developed and implemented procedure manuals for the IACUC and IBC, respectively.
3. Performed pre-review of all IACUC and IBC protocols.
4. Moved IBC review process from paper-based system to semi-electronically system (emails, PDFs)
5. Developed specification for fully electronic web-based IBC review (based on IACUC processes which was already a web-based system; BRAIN); specification were implemented in 2015.
6. Participated in all IACUC animal facility inspections and IACUC/IBC investigations of non-compliance concerns.
7. Organized and participated in semi-annual program review of the animal program.
8. Participated in 2 AALAC site visit.
9. Developed training for IACUC and IBC members as well as
10. Annually taught one lecture on ethical use of animals in research to medical and graduate students.

1999 – 2003
40 hours/week

Instructor
Breast Center, Department of Medicine, Baylor College of Medicine,
Houston, Texas

Responsibilities:

1. Developed an independent research program
2. Prepared and submitted research proposals to federal and private funding agencies.
3. Designed and performed experiments in cancer biology using state-of-the-art molecular and cell biological laboratory techniques.
4. Trained and supervised laboratory technicians.
5. Prepared and submitted manuscripts for publications in scientific journals.
6. Presented research findings at national and international meetings.
7. Taught undergraduate and graduate students.

1996 – 1999

Postdoctoral Research Fellow
Laboratory of Dr. Suzanne A.W. Fuqua
Department of Medicine/Oncology, University of Texas / Health
Science Center at San Antonio, Texas

Responsibilities:

1. Designed and performed experiments in cancer biology using state-of-the-art molecular and cell biological laboratory techniques.
2. Developed transgenic mouse lines over-expressing wild-type estrogen receptor and mutant estrogen receptor.
3. Collected, compiled and analyzed research data.
4. Prepared and submitted manuscripts for publications in scientific journals.
5. Presented research findings at national and international meetings.

1994 – 1996

Postdoctoral Research Fellow
Laboratory of Dr. Eileen Lafer
Center for Molecular Medicine / Institute of Biotechnology, University
of Texas / Health Science Center at San Antonio, Texas

Responsibilities:

1. Designed and performed experiments in neurobiology using state-of-the-art molecular and biochemical laboratory techniques.
2. Collected, compiled and analyzed research data.
3. Presented research findings at national and international meetings.

1991 – 1994

Graduate Research Assistant
Laboratory of Professor Peter K. Mueller

Institute of Medical Molecular Biology, Medical University of
Luebeck, Germany
Ph.D. Thesis: Characterization of Vigilin”

PROFESSIONAL MEMBERSHIP

2015 – present Midwest Area Biosafety Network
2010 – present American Biological Safety Association (serves on the membership
committee)
2003 – 2010 Applied Research Ethics National Association (ARENA)
2004 – 2008 American Association for Laboratory Animal Science (AALAS)
2001 – 2003 Endocrine Society
2000 – 2003 American Association for Cancer Research

HONORS AND AWARDS

2002 – 2003 SPORE Career Development Award

OTHER

01/2017 Registered Biosafety Professional
2010 – 2015 Member of the Institutional Biosafety Committee at Rice University
2006 – 2015 Member of the Texas Children’s Cancer Center Protocol Review
Committee
10/2007-12/2012 Certified Professional IACUC Administrator

PUBLICATIONS

Herynk, M.H., Hopp, T., Cui, Y., Niu, A., Corona-Rodriguez, A., and Fuqua, S.A.W.: A hypersensitive estrogen receptor a mutation that alters dynamic protein interactions. *Breast Cancer Res Treat.* 2010 Jul; 122(2):381-93. PMID: 19842032

Herynk MH, Lewis MT, Hopp TA, Medina D, Corona-Rodriguez A, Cui Y, Beyer AR, Fuqua SA. Accelerated mammary maturation and differentiation, and delayed MMTVneu-induced tumorigenesis of K303R mutant ERalpha transgenic mice. *Oncogene.* 2009 Jun 29. [Epub ahead of print]

Husser RC and Hopp TA. Response to Protocol Review Scenario: Taking protocols for granted. *Lab Anim (NY).* 2006 Sep;35(8):16.

Martin MD, Hilsenbeck SG, Mohsin SK, Hopp TA, Clark GM, Osborne CK, Allred DC, and O'connell P. Breast tumors that overexpress nuclear metastasis-associated 1 (MTA1) protein

have high recurrence risks but enhanced responses to systemic therapies. *Breast Cancer Res Treat.* 2005 Oct 22;:1-6 [Epub ahead of print]

Hopp, TA. Becoming an IACUC Administrator: Why would a scientist leave laboratory research for a career as an IACUC administrator? *Contemporary Topics* 63 (6): 74-76, 2004.

Hopp TA, Weiss HL, Parra IS, Cui Y, Osborne CK, Fuqua SA. Low Levels of Estrogen Receptor {beta} Protein Predict Resistance to Tamoxifen Therapy in Breast Cancer. *Clin Cancer Res.* 10(22):7490-7499, 2004.

Silverman J, Kuhlman SM, Hill LR, Hopp T, and Josten M. Pig surgery: can reduction be a protocol violation? *Lab Anim (NY)* 33(6):14, 2004.

deGraffenried LA, Hopp TA, Valente AJ, Clark RA, and Fuqua SA. Regulation of the estrogen receptor alpha minimal promoter by Sp1, USF-1 and ERalpha. *Breast Cancer Res Treat* 85(2):111-20, 2004.

Hopp TA, Weiss HL, Hilsenbeck SG, Cui Y, Allred DC, Horwitz KB, and Fuqua SA. Breast cancer patients with progesterone receptor PR-A-Rich tumors have poorer disease-free survival rates. *Clin Cancer Res* 10(8):2751-60, 2004.

Cui X, Lazard Z, Zhang P, Hopp TA, and Lee AV. Progesterone crosstalks with insulin-like growth factor signaling in breast cancer cells via induction of insulin receptor substrate-2. *Oncogene* 22(44):6937-41, 2003.

Osborne CK, Bardou V, Hopp TA, Chamness GC, Hilsenbeck SG, Fuqua SA, Wong J, Allred DC, Clark GM, and Schiff R. Role of the estrogen receptor coactivator AIB1 (SRC-3) and HER-2/neu in tamoxifen resistance in breast cancer. *J Natl Cancer Inst* 95(5):353-61, 2003.

Fu M, Wang C, Wang J, Zhang X, Sakamaki T, Yeung YG, Chang C, Hopp T, Fuqua SA, Jaffray E, Hay RT, Palvimo JJ, Janne OA, and Pestell RG. Androgen receptor acetylation governs trans activation and MEKK1-induced apoptosis without affecting in vitro sumoylation and trans-repression function. *Mol Cell Biol* 22(10):3373-88, 2002.

Wang C, Fu M, Angeletti RH, Siconolfi-Baez L, Reutens AT, Albanese C, Lisanti MP, Katzenellenbogen BS, Kato S, Hopp T, Fuqua SA, Lopez GN, Kushner PJ, and Pestell RG. Direct acetylation of the estrogen receptor alpha hinge region by p300 regulates transactivation and hormone sensitivity. *J Biol Chem* 276(21):18375-83, 2001.

Zhou Q, Hopp T, Fuqua SA, and Steeg PS. Cyclin D1 in breast premalignancy and early breast cancer: implications for prevention and treatment. *Cancer Lett* 162(1):3-17, 2001.

Fuqua SA, Wiltschke C, Zhang QX, Borg A, Castles CG, Friedrichs WE, Hopp T, Hilsenbeck S, Mohsin S, O'Connell P, and Allred DC. A hypersensitive estrogen receptor-alpha mutation in premalignant breast lesions. *Cancer Res* 60(15):4026-9, 2000.

Oesterreich S, Zhang Q, Hopp T, Fuqua SA, Michaelis M, Zhao HH, Davie JR, Osborne CK, and Lee AV. Tamoxifen-bound estrogen receptor (ER) strongly interacts with the nuclear matrix protein HET/SAF-B, a novel inhibitor of ER-mediated transactivation. *Mol Endocrinol*. 14(3):369-81, 2000.

Hopp TA and Fuqua SA. Estrogen receptor variants. *J Mammary Gland Biol Neoplasia* 3(1):73-83, 1998.

Neu-Yilik G, Hopp-Christensen TA, Dobberstein B, and Muller PK. Vigilin is associated with free and membrane bound ribosomes. *Eur J Cell Biol* 40(63):84, 1994

Neu-Yilik G, Zorbas H, Gloe TR, Raabe HM, Hopp-Christensen TA, and Muller PK. Vigilin is a cytoplasmic protein. A study on its expression in primary cells and in established cell lines of different species. *Eur J Biochem*. 213(2):727-36, 1993

BOOK CHAPTERS

Hopp TA and Fuqua SAW. Estrogen and progesterone receptor in breast cancer. In "Encyclopedia of Hormones" (Editors: Henry HL and Norman AW), Elsevier Science, San Diego, CA: 573-578, 2003

Hopp TA and Fuqua SAW. Signalling by steroid receptors. In "The Cancer Handbook" (Editor: Alison MR), Vol 1: 135-147, 2002.

Hopp TA and Fuqua SA. Steroid and nuclear receptor polymorphism resulting in hormone resistance and hormone independence. In "Genetic Polymorphism and Susceptibility to Disease": 109-138, 2001

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Previous Supervisor:
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