

Protocol - ISOLATING DNA FROM MOUSE TAILS

- 1) Pipette 475ul Jim's tail buffer into a tube
- 2) Add 25ul or .025ml protease K
- 3) Place in 55 degrees water bath OVERNIGHT
- 4) Add 170ul of saturated NaCl solution
- 5) Shake and centrifuge for 10 minutes @ 14000g
- 6) Label one set of microtubes
- 7) Remove the supernant (transparent) part of the centrifuged solution into labeled tube
- 8) Fill the supernant with 100% ethanol, shake well. (NOTE: The DNA should suspend in solution)
- 9) Centrifuge for 10 minutes @ 14000g
- 10) Carefully vacuum the supernant from the tube, and fill tube with 800ul of 80% alcohol.
- 11) Centrifuged for 10 minutes at 14000g
- 12) Use Vacuum apparatus to extract the top of clear alcohol solution from the centrifuge tube, leaving the DNA precipitant. Let stand for 10 minutes to allow the remaining alcohol to evaporate from the tube.
- 13) Pipette 300ul (or 150ul if DNA is not visible) of TE solution into the dry DNA tube and let stand overnight.